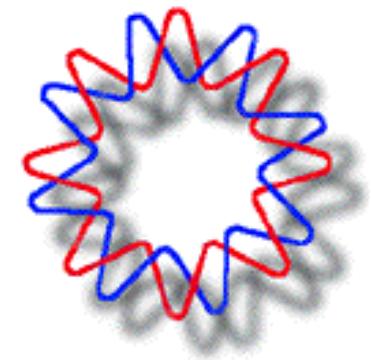


Producción de etanol carburante a partir de rastrojo de maíz con bacterias etanologénicas

**Fuel ethanol production from corn stover
with ethanogenic bacteria**



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*Manizales, Col.
30/May/2017*

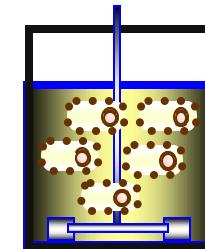
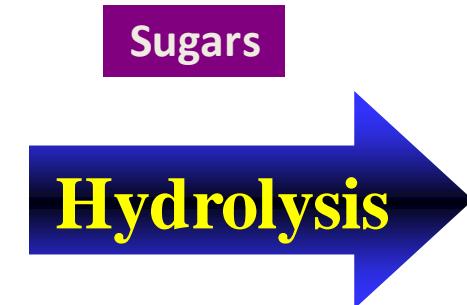
Generation Ethanol (Agro-Fuels) and (Agro-) Chemicals from Lignocellulose



Artificial
CO₂ cycle



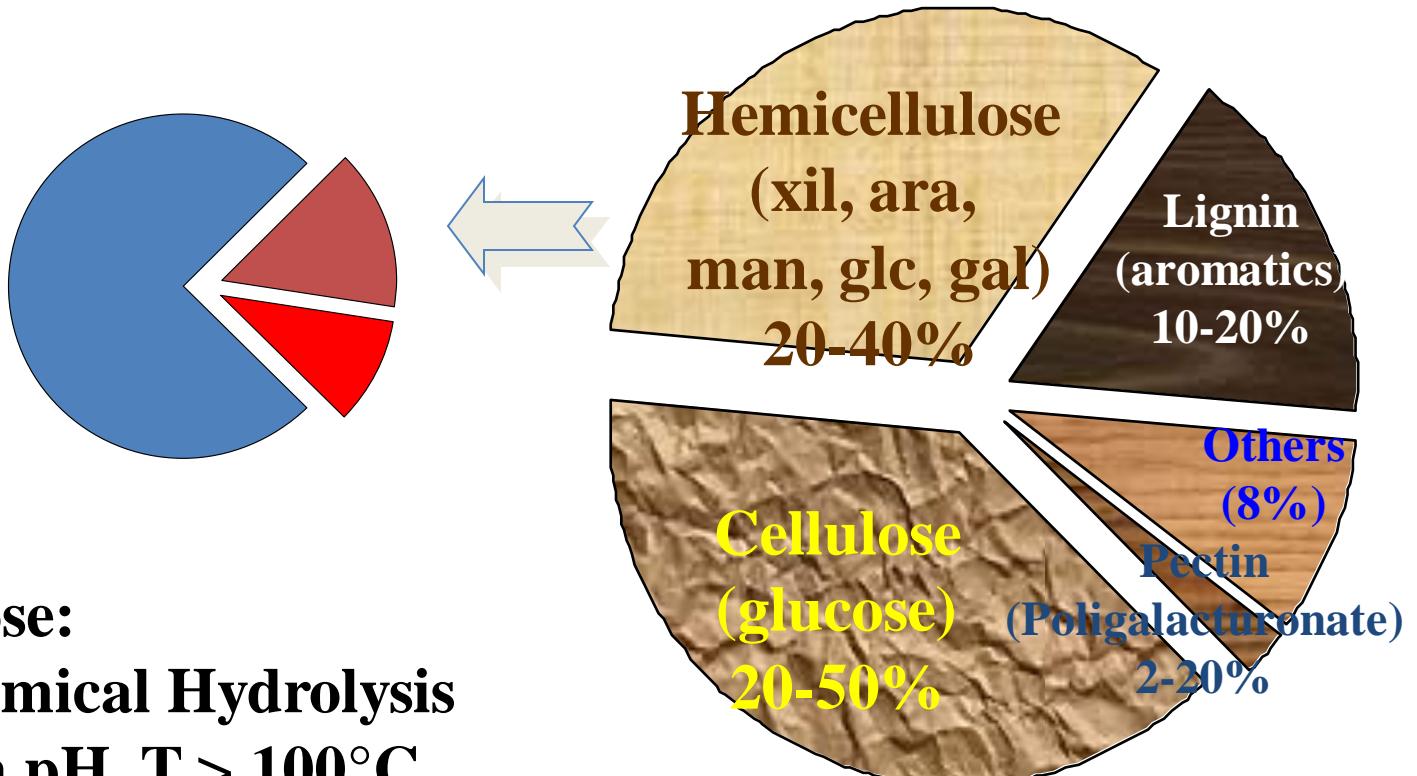
Chemicals
Ethanol (Fuel)



Fermentation

Purpose: Design microorganism and process to transform ALL the SUGARS contained into lignocellulose (cellulose: glucose & hemicellulose: pentoses, hexoses, disaccharides) to ethanol (or other chemicals)

Lignocellulose



Hemicellulose:
Thermochemical Hydrolysis
Low or high pH, T > 100°C

Cellulose:
Enzymatic Hydrolysis
pH 4.5-5, 50°C or higher

Pentoses + Hexoses
(Xil + Ara) + (Gluc+Man+Gal)
+ Acetate + Glucuronic Acid
+ Toxins

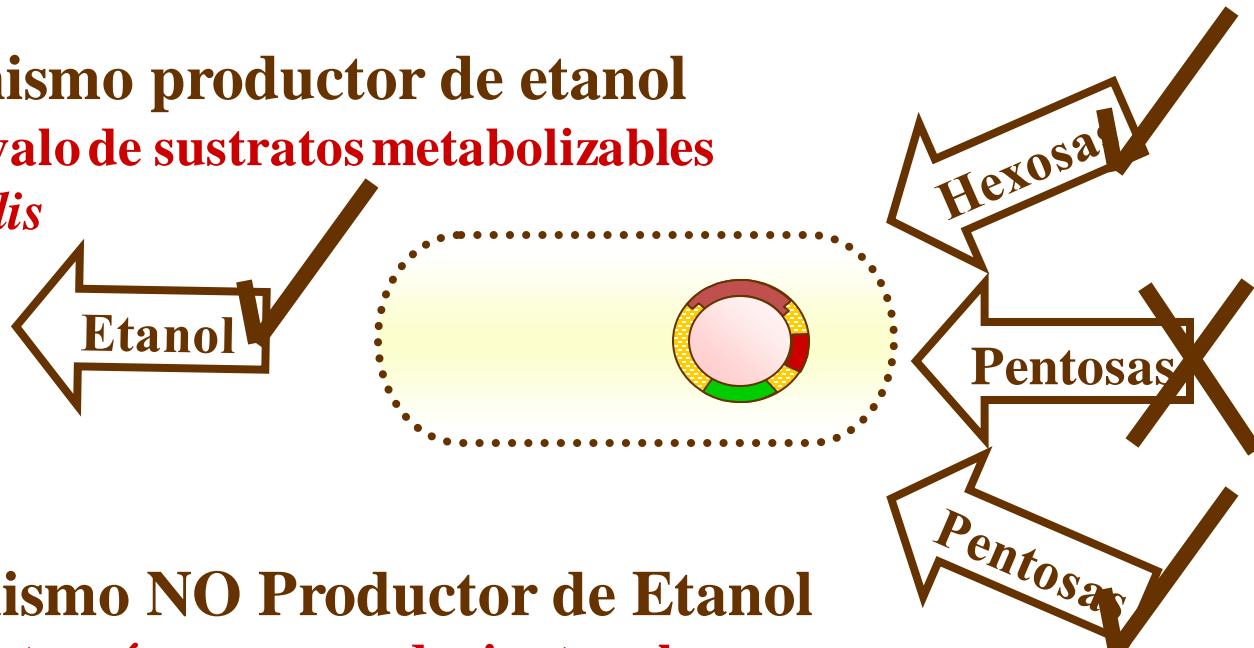
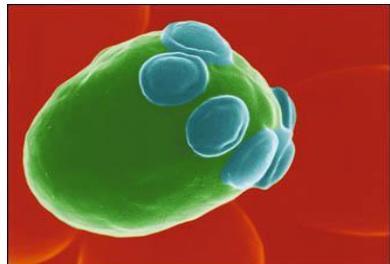
Dos principales estrategias: Pentosas → Etanol

Ingeniería de Vías Metabólicas

A: Microorganismo productor de etanol

Extender el intervalo de sustratos metabolizables

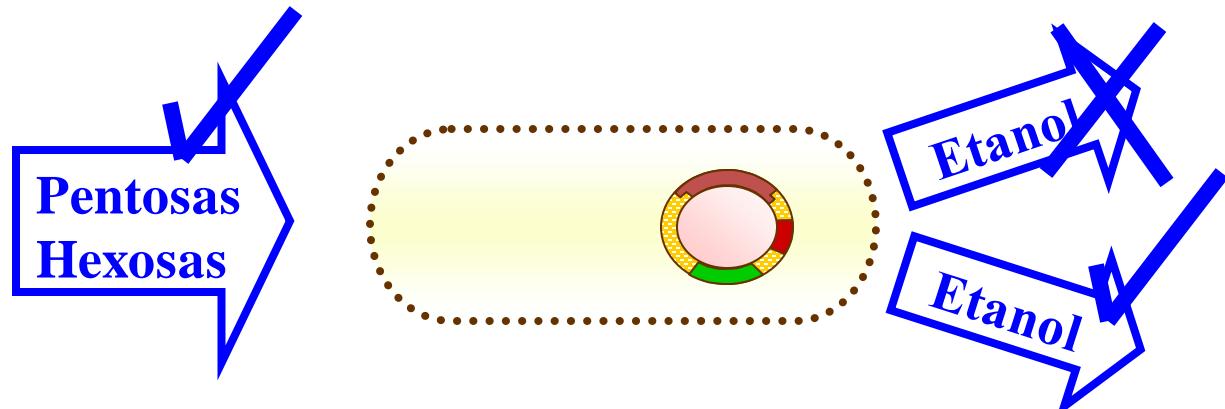
Zymomonas mobilis



B: Microorganismo NO Productor de Etanol

Complementar vías para producir etanol

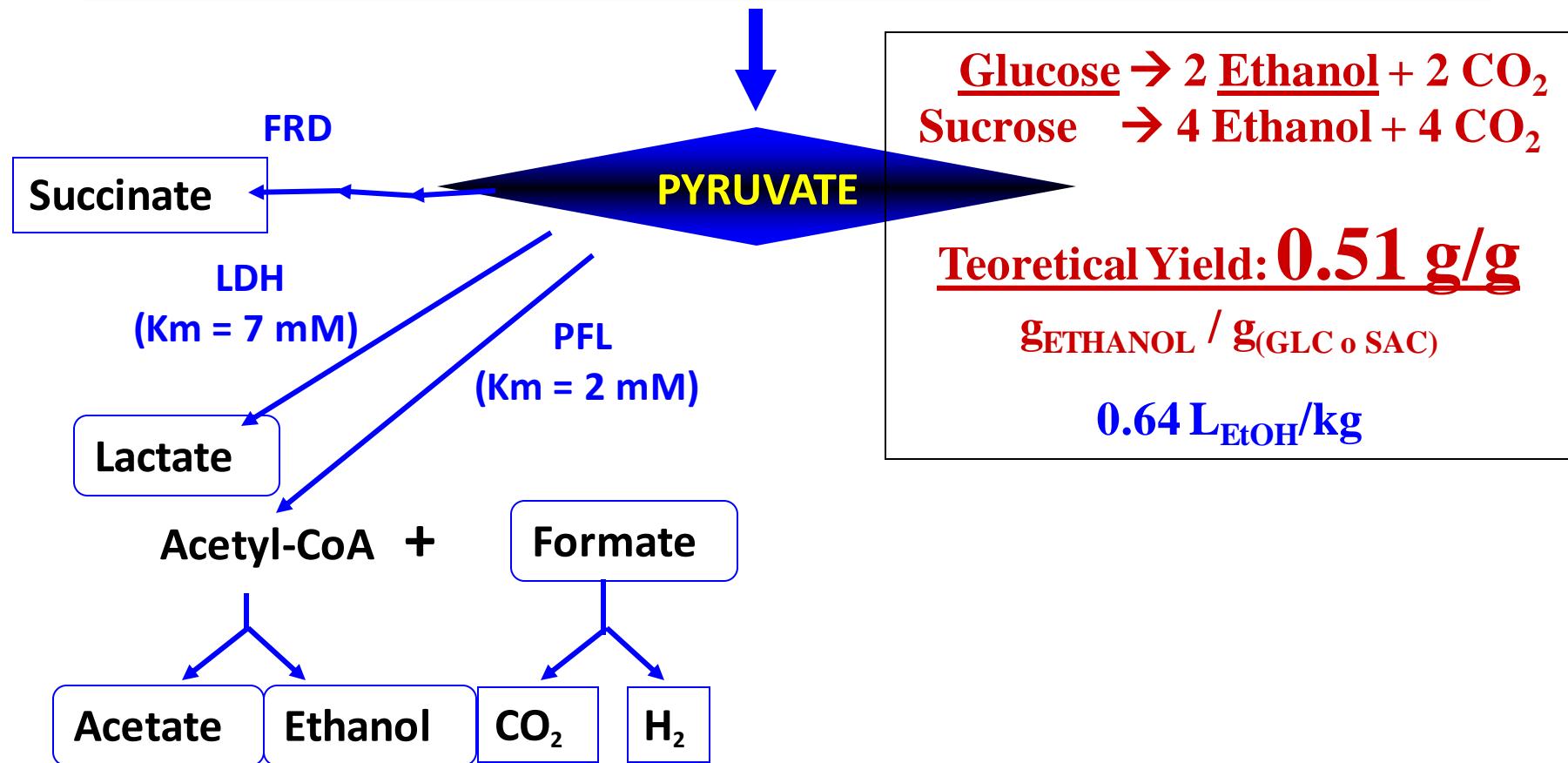
Escherichia coli



Escherichia coli Fermentative Phatways

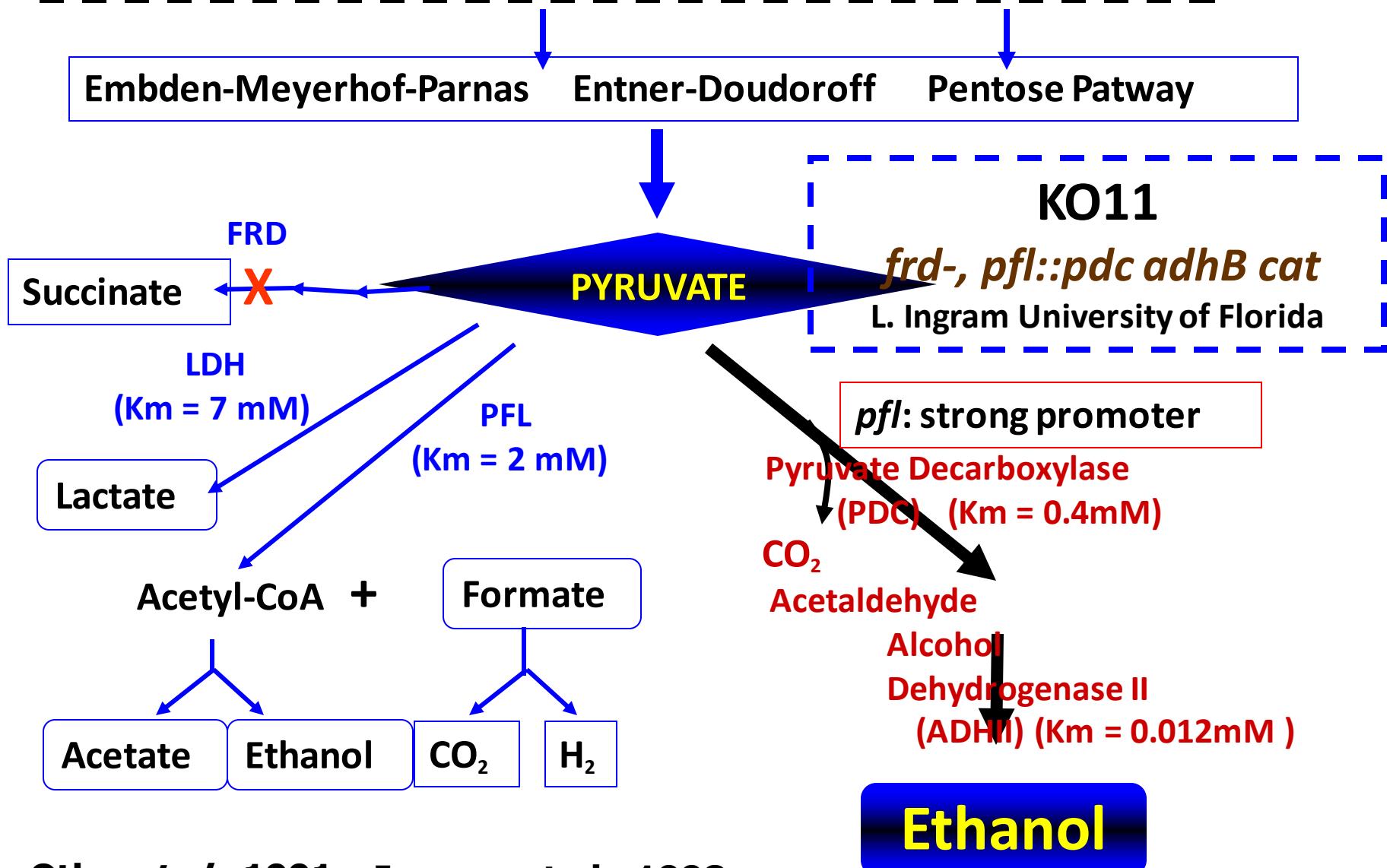
HEXOSES (Glc, Fru, Gal, Man etc.) + PENTOSES (Xyl, Ara, Rib, Xylu, etc.)

Embden-Meyerhof-Parnas Entner-Doudoroff Pentose Patway



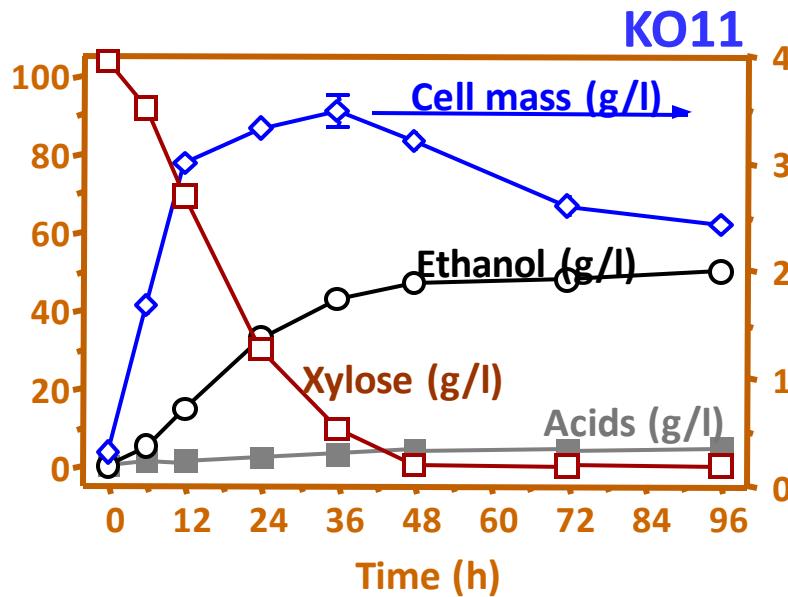
Escherichia coli Etanologénica: 1ra Generación

HEXOSES (Glc, Fru, Gal, Man etc.) + PENTOSES (Xyl, Ara, Rib, Xylu, etc.)

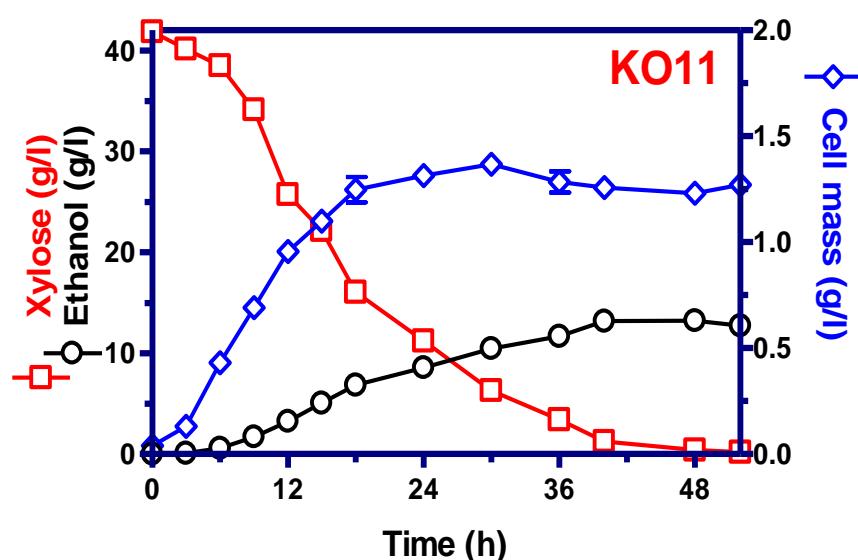


Otha *et al.*, 1991 Ingram *et al.*, 1998

KO11: Xyl (10%) Rich Medium



Xyl (4%): Mineral Media



In comparison with rich media, in mineral media there are reductions by:
 57_{Glc} & $63_{\text{Xyl}}\%$ in cell mass formation; 25% in the specific growth rate
 70_{Glc} & $60_{\text{Xyl}}\%$ in the specific sugar consumption rate

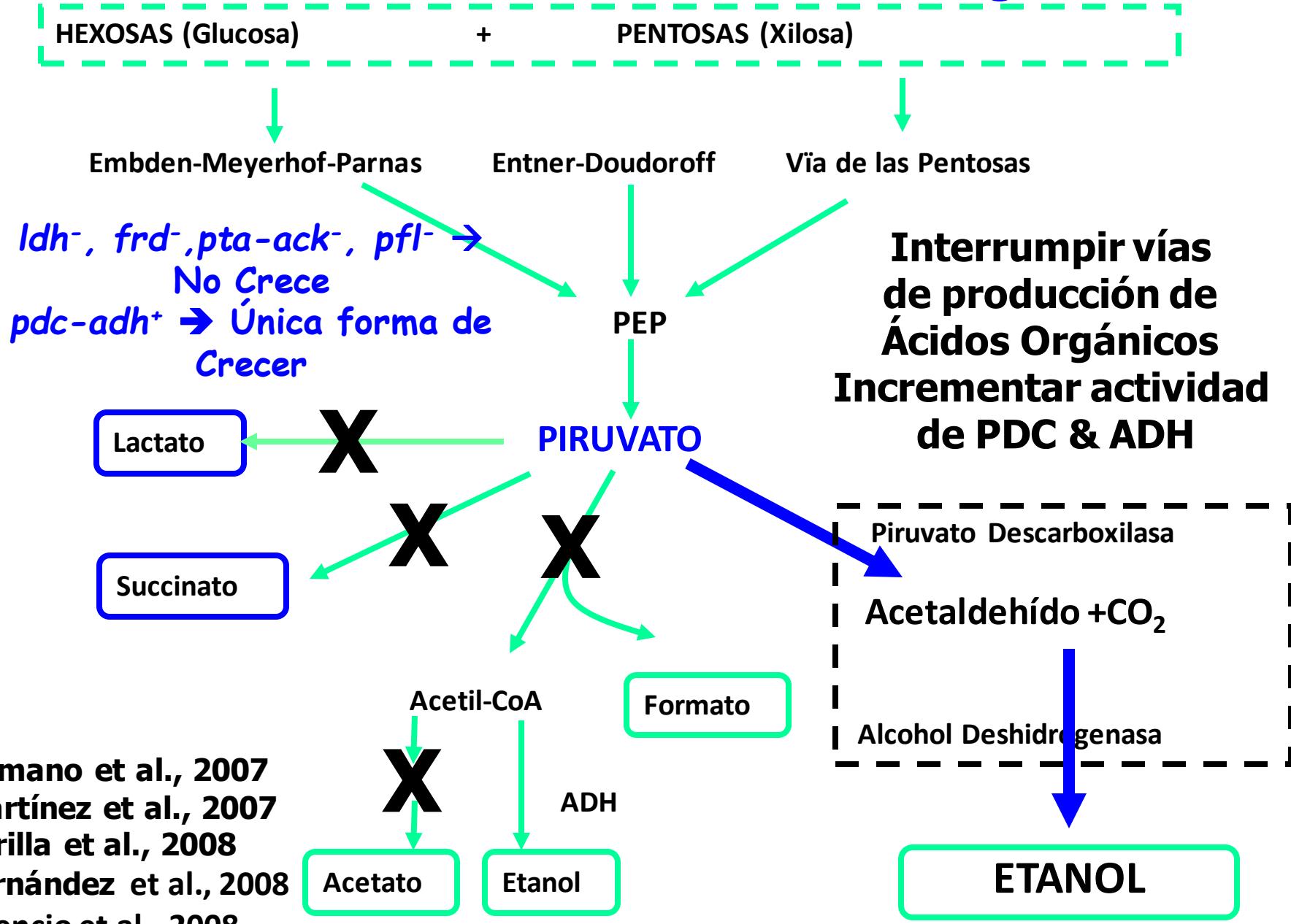
And Q_{EtOH} is reduced to 0.42_{Glc} & $0.33_{\text{Xyl}} \text{ g}_{\text{Et-OH}}/\text{l h}$, for glucose and xylose, respectively

37°C, 100 rpm, pH 7.0

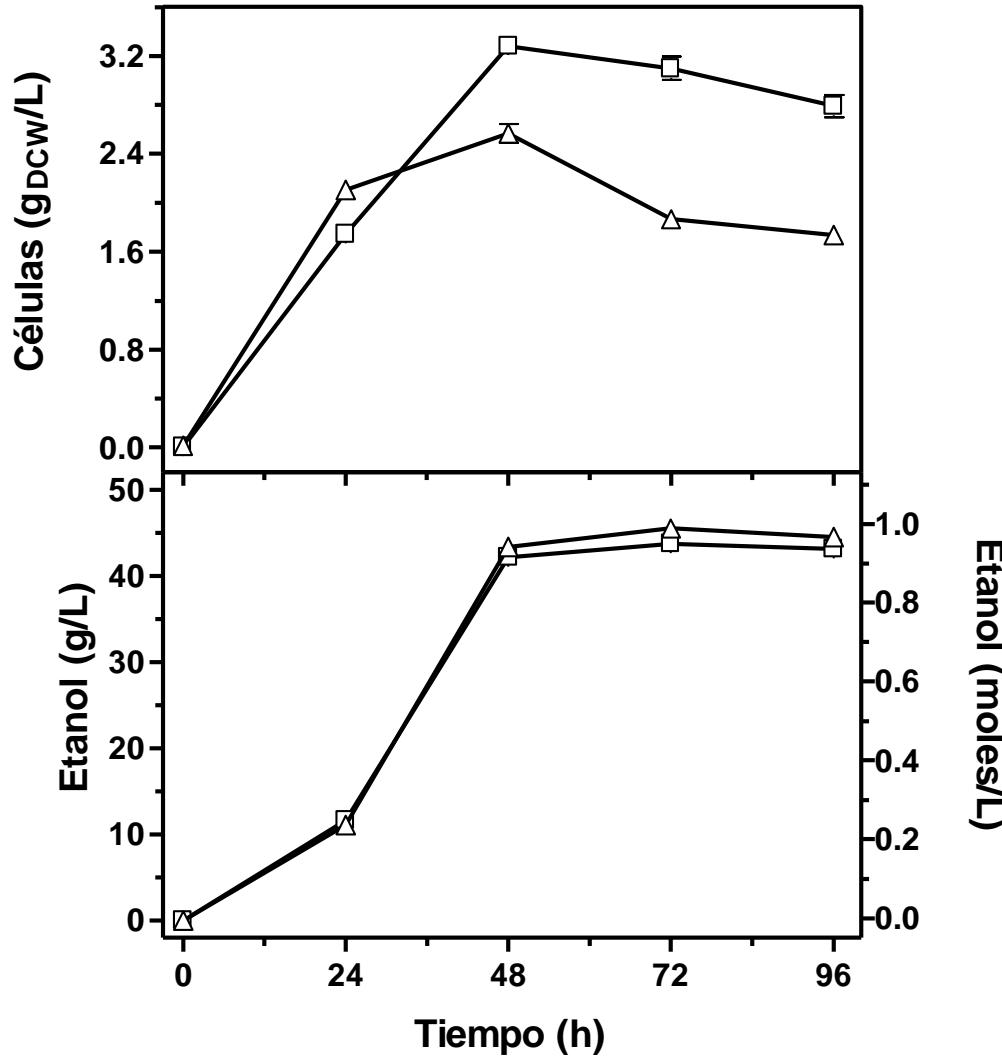
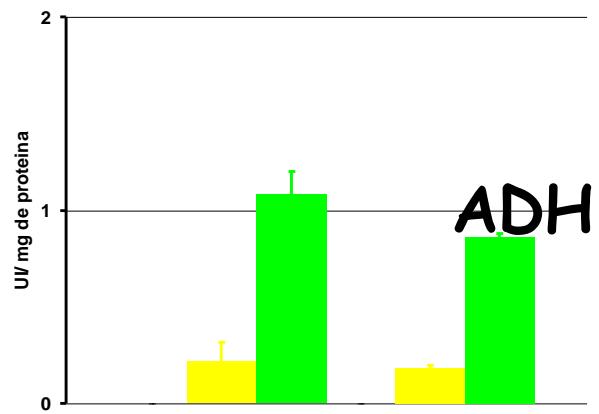
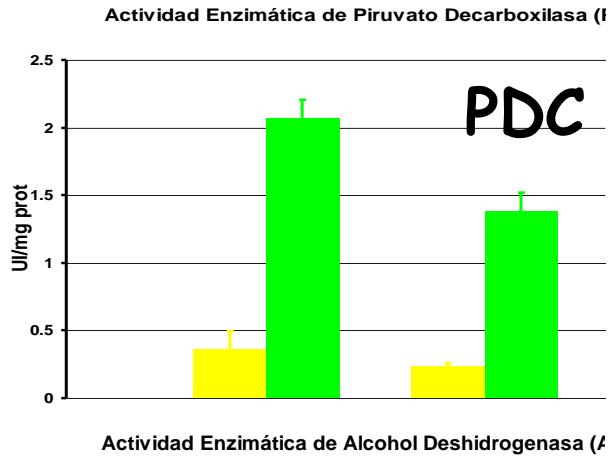
Yield >95%

Yield 60%

2da Generación de *E. coli* Etanologénicas



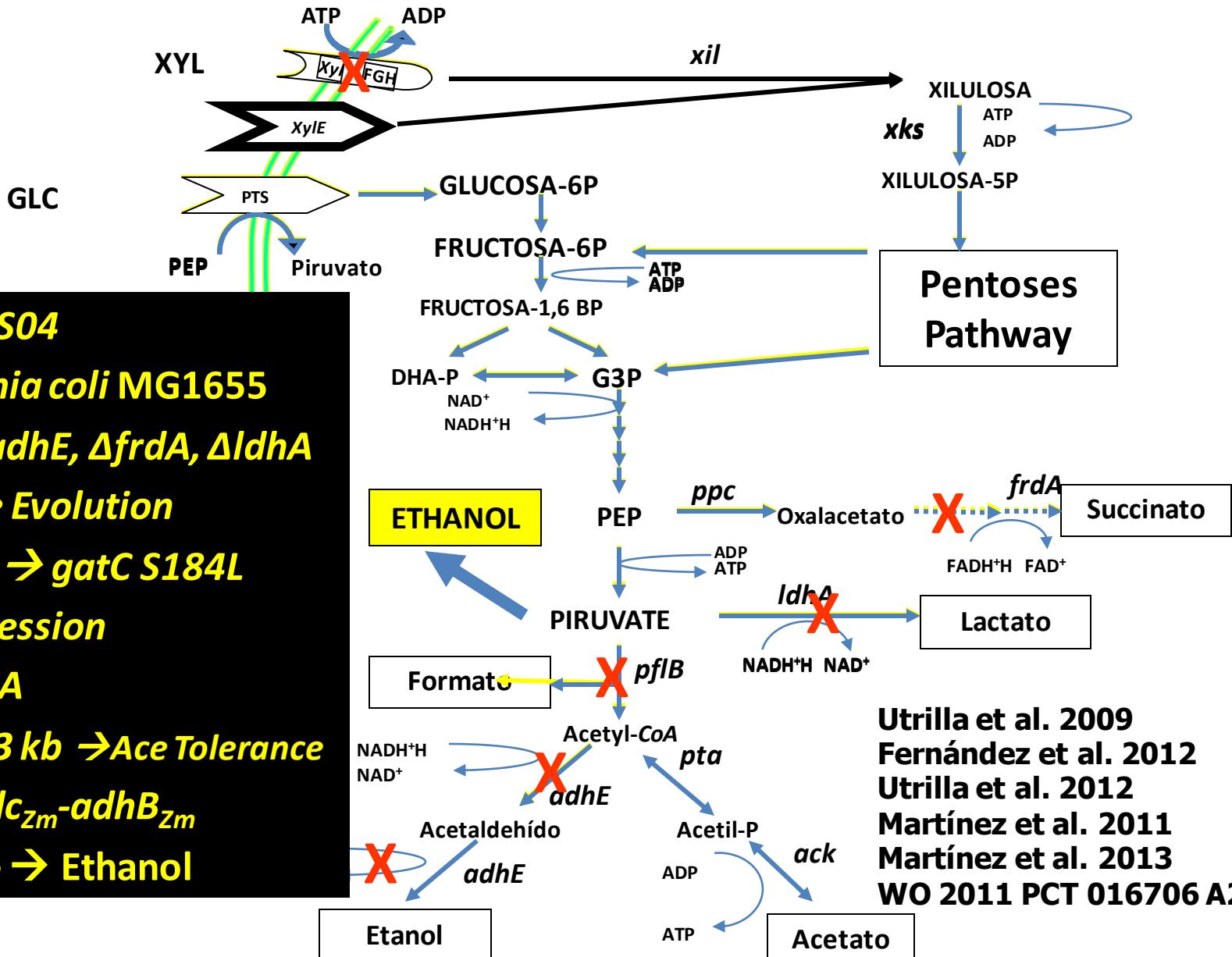
Ethanol Mineral Media: Xyl-Glc



Yield >90%

Ethanologenic *E. coli* strain to use pentose-hexose mixtures

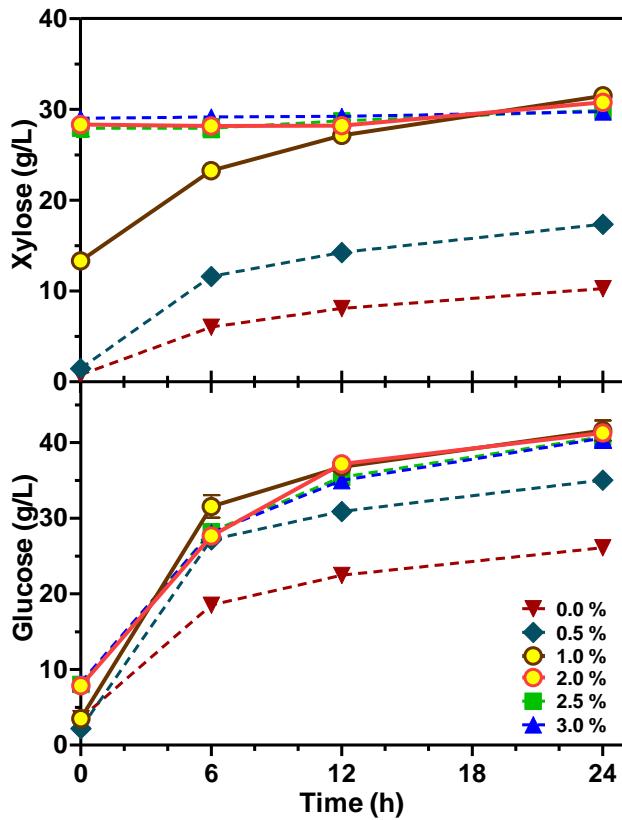
MG1655: $\Delta pflB$, $\Delta adhE$, $\Delta frdA$, $\Delta xylFGH$, Δldh , $PpflB::pdc-adh_{Zm}$



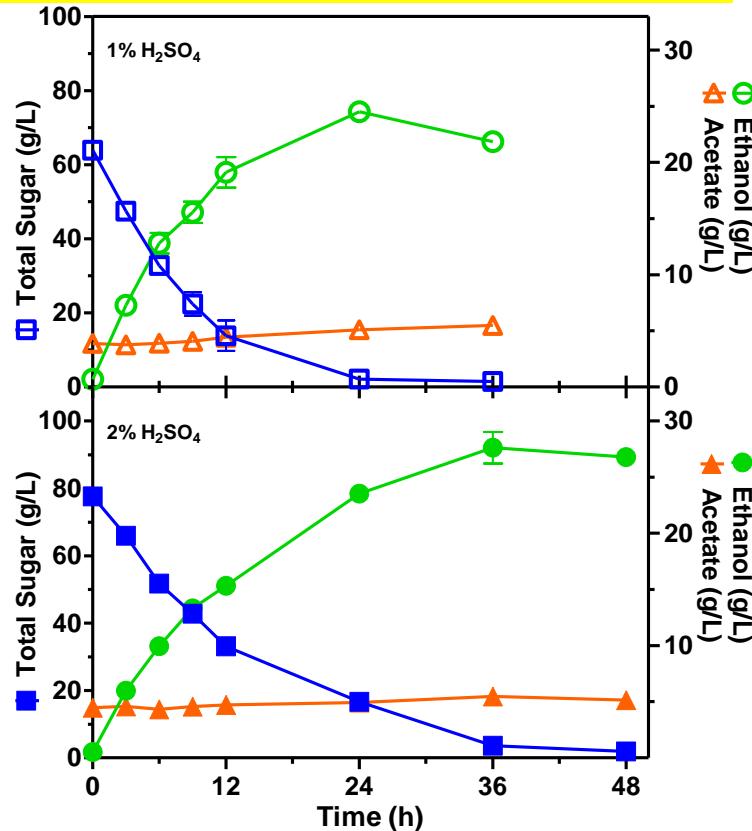
- Strain MS04
- Escherichia coli MG1655
- $\Delta pflB, \Delta adhE, \Delta frdA, \Delta ldhA$
- Adaptive Evolution
- $\Delta xylFGH \rightarrow gatC S184L$
- Pdh expression
- $\Delta midarpA$
- $\Delta reg 27.3\text{ kb} \rightarrow Ace$ Tolerance
- $PpflB::pdc_{Zm}-adhB_{Zm}$
- Pyruvate → Ethanol

Stover from White Corn: Sequential: Thermochemical Hydrolysis, Enzymatic Saccharification and Fermentation

Diluted Acid Pretreated Slurries
 Saccharification Peg Mixer Reactor
 15 UPF/gBA, 30 UCB/gBA
 50 °C pH = 5.0, 60 rpm



Non-aerated Cultures with Ethanologenic *E. coli* MS04, 3.7 g/L, 0.2 L, 37°C, pH 7, 100 rpm.
No salts were added. No detox.



Most of the Xyl & Glc are released for pretreated S-WT above 1% SA in 12 h

All sugars are fermented to ethanol by ethanologenic *E. coli* MS04 in 36 h

Corn and Agaves



% DM (w/w)	Corn Stover	Agave Bagasse
Glucan	29.9 (1.2)	38.6 (2.5)
Xylan	19.6 (0.8)	13.6 (1.1)
Arabinan	3.4 (0.2)	3.4 (0.1)
Acetyl	4.9 (0.4)	4.2 (0.9)
Lignin	21.7 (0.6)	16.0 (1.3)
Extractives	6.8 (0.2)	12.9 (0.7)
Ashes	10.9 (0.7)	5.5 (0.4)
Other	2.8 (2.05)	4.1 (0.5)

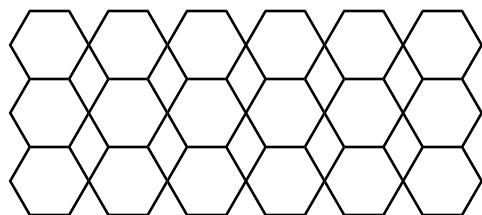
**275 L_{EtOH}/Ton Dry Corn Stover
Bajio 2012: 3,000,000 Tons CS →
825 x 10⁶ Liters**

**330 L_{EtOH}/Ton Dry Agave Bagasse
Tequila 2008: 220,000 Tons AB →
72 x 10⁶ Liters**

Consolidated Bioprocessing

Hydrolases:

cellulases and
 β -glucosidases



Glucose



EtOH

Pentoses:
xylose and
arabinose

Hexoses:
glucose,
mannose and
galactose

van Zyl et al. (2007) Adv Biochem Eng Biotechnol.
Lynd et al. (2005) Curr Opin Biotechnol.



E. coli MS04:
Homo-Ethanogenic strain
Hexoses Pentoses
Acetate tolerant



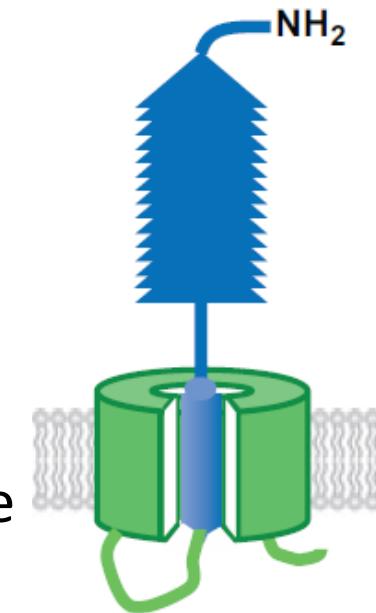
Purpose:
MS04 derivative strain with the ability to secrete a β -glucosidases to produce ethanol from cellobiose → Cellulose At relatively high Temperature

Type Va Secretion System (Autotransporter pathway)

Translocation Unit
(TU)



External
Membrane



Dautin and Bernstein (2007) Annu Rev Microbiol

Autotransporter	Source
AIDA-I of diarrheagenic <i>E. coli</i> has been used for the secretion of many heterologous proteins.	Maurer et al. (1997) J Bacteriol. Jose and Meyer (2007) Microbiol Mol Biol Rev.

Beta-glucosidase to be secreted in *E. coli*

Parameter	<i>T. fusca</i> BglC ^b	<i>A. Niger</i> BG _S ^c
Temperature (°C)	50	65
pH	7	3.5-4
MW (kDa)	53	200
Vmax (mmol min ⁻¹ mg ⁻¹) ^a	28	68
Km (mM) ^a	0.35	2.7

^a Parameters obtained at 25°C.

^b Spiridonov and Wilson (2001) Curr Microbiol.

^c Seidle et al. (2004) Protein J.

MS04/pAIDABglCRHis

Cell surface display of the β -glucosidase BgIC from *Thermobifida fusca* on the surface of the ethanologenic *Escherichia coli* strain MS04
For ethanol production from cellobiose

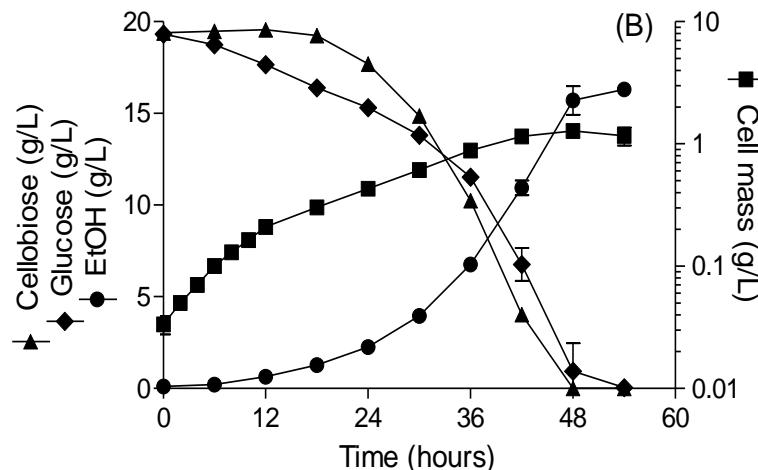
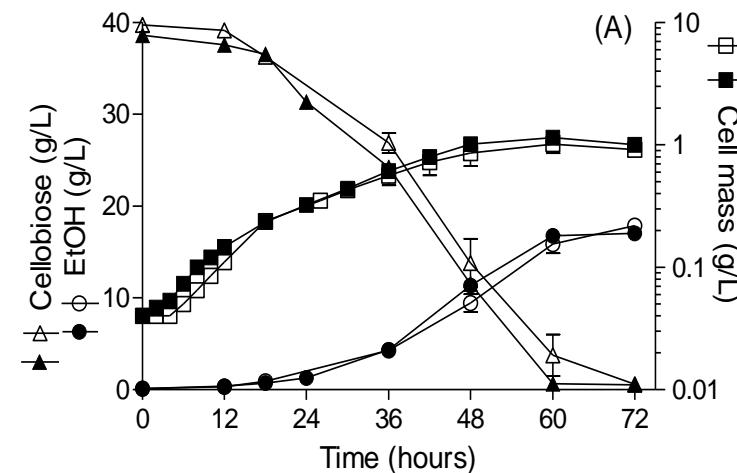
Cellobiose

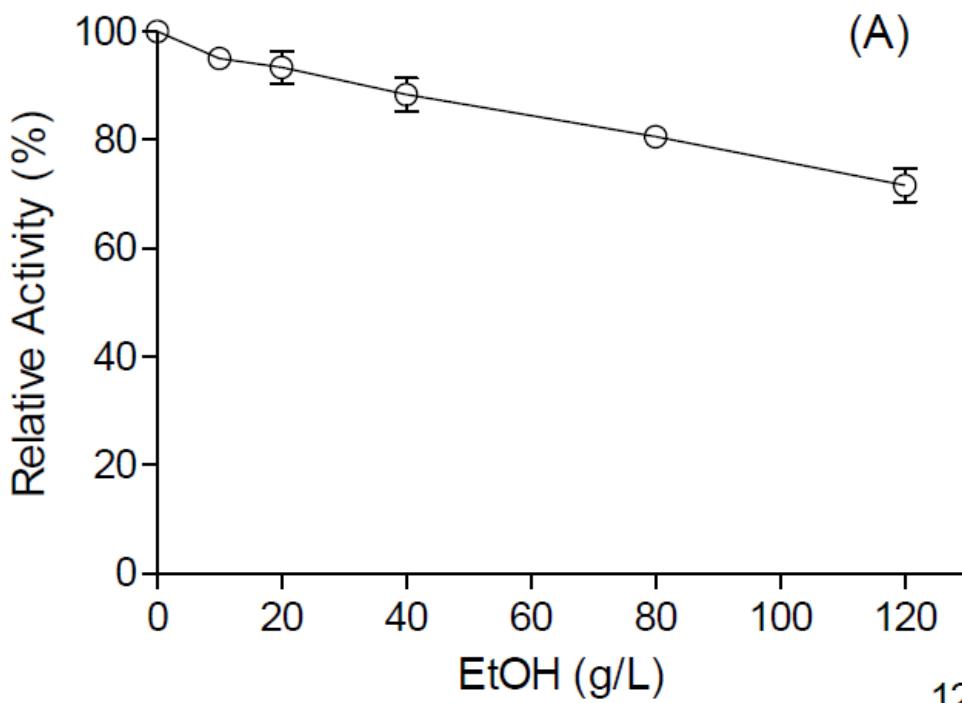
β -glucosidasa *Thermobifida fusca*: BgIC

T. (°C)	pH	MW (kDa)
50	7	53

Cellobiose – Glucose

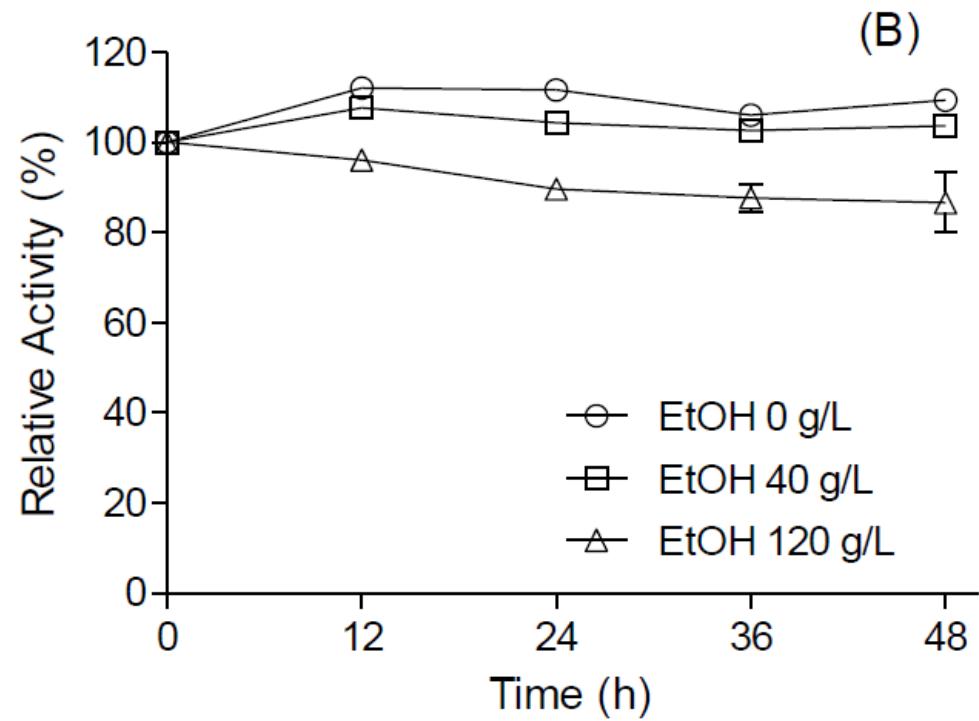
Muñoz-Gutiérrez et al., 2012





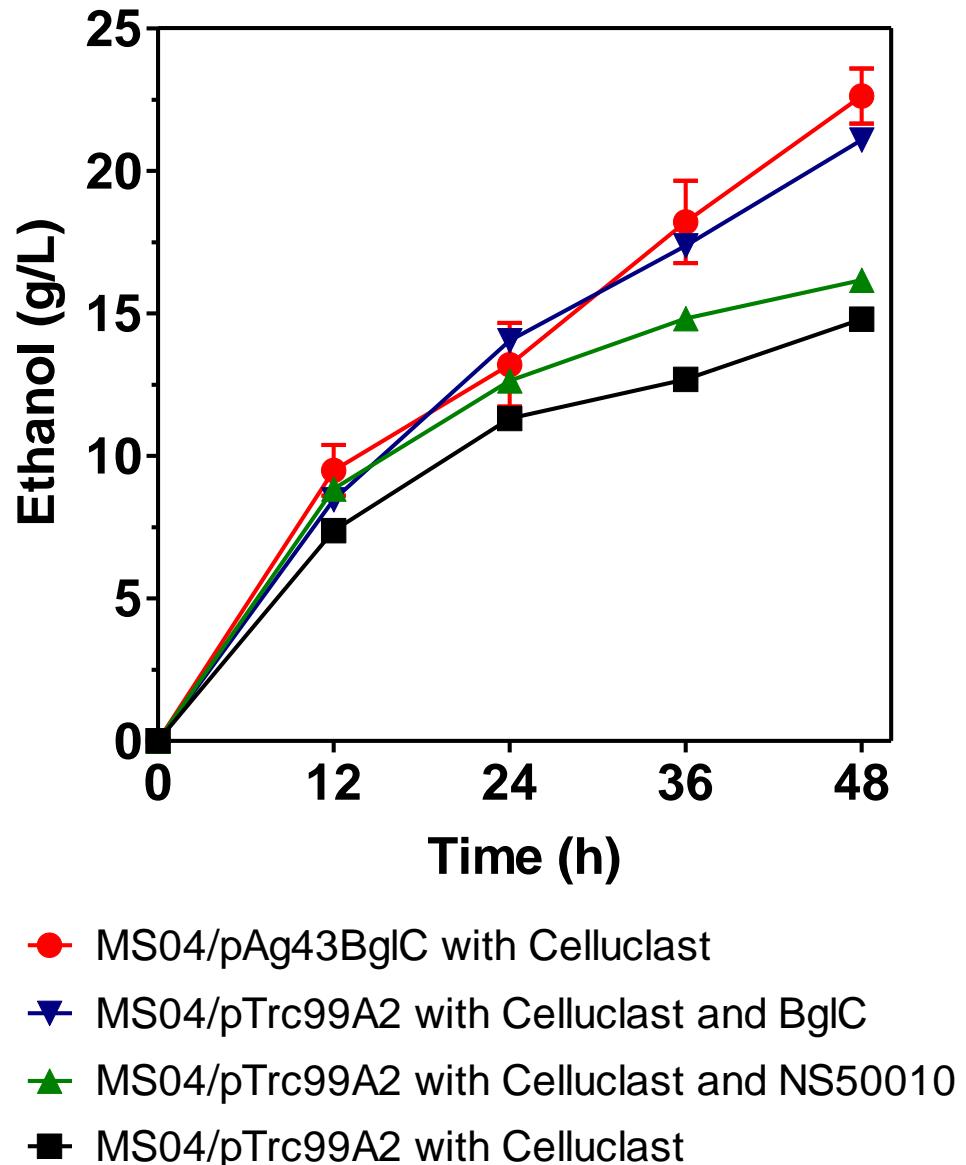
Effect of EtOH on the
stability of BglC attached to
MS04

Effect of EtOH on the
activity of BglC attached to
MS04



SSF AVICEL 45°C pH 6

- Ethanol production during the SSF process of 40 g/L Avicel by MS04 carrying plasmid pAg43BglC or pTrc99A2.
- With Exo-endo Cellulase addition



Remarks. From Cellulose. Other applications.

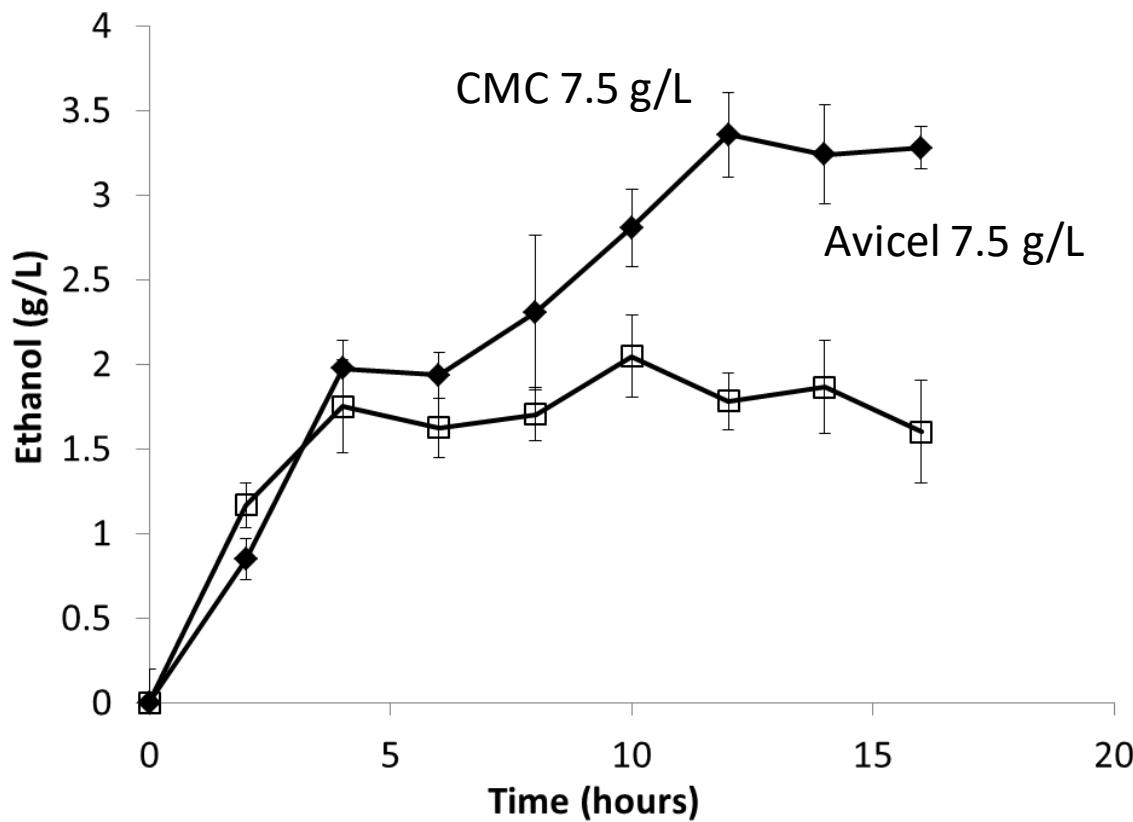
MSO4 – EndoG

Ethanol production

MSO4-EndoG / MSO4 –
BglC

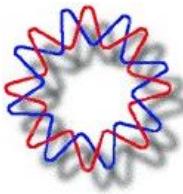
No cellulase addition.
45°C, pH 6

Loaces, I, et al., 2016





Gracias



- ♦ **CONACyT**
- ♦ **UNAM PAPIIT – DGAPA**

- ♦ **Estudiantes y colegas**
- ♦ **Instituto de
Biotecnología - UNAM**